One-Pot Synthesis of Esters of Cyclopropane Carboxylic Acids via Tandem Vicarious Nucleophilic Substitution—Michael Addition Process

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Supporting Information

ABSTRACT: α -Chlorocarbanions generated via base-induced vicarious nucleophilic substitution reaction of alkyl dichloroacetates with nitroarenes react with Michael acceptors to give esters of cyclopropane carboxylic acids substituted with *p*nitroaromatic rings.

INTRODUCTION

A cyclopropane ring is present in many biologically active compounds and pharmaceuticals.¹ Substituted cyclopropanes are valuable intermediates in the synthesis of larger carbo- and heterocyclic rings via three-membered ring opening, followed by cycloaddition.² Because of that, synthesis of substituted cyclopropanes is of substantial interest and is the subject of many publications.³ In general, there are two major ways of construction of cyclopropane rings: addition of carbenes and carbenoids to C=C double bonds and intramolecular 1,3substitution in carbanions containing nucleofugal groups in γ positions. The second pathway includes generation of γ halocarbanions via addition of α -halocarbanions to Michael acceptors, deprotonation of γ -haloalkyl sulfones, esters and nitriles of γ -haloalkanoic acids, alkylation of methylenic carbanions with 1,2-dihaloalkanes, and reactions of Michael acceptors with ylides. Because of their instability, the reactions of α -halocarbanions with Michael acceptors are usually carried out via deprotonation of α -haloalkenoic nitriles, esters, sulfones, etc., in the presence of the Michael acceptors.

In this paper, we wish to report the synthesis of substituted cyclopropanes in reaction of Michael acceptors with α -chlorocarbanions generated via vicarious nucleophilic substitution (VNS) of hydrogen in nitroarenes.

VNS is a general reaction between α -halocarbanions and nitroarenes that proceeds via addition of the carbanions to the electron-deficient nitroaromatic rings in positions occupied by hydrogen to form σ^{H} adducts. Further base-induced β elimination of hydrogen halide results in formation of nitrobenzylic carbanions of the products. Protonation of these carbanions gives products of replacement of the ring hydrogen with the carbanion moiety (Scheme 1).⁴

Besides of protonation to give products of VNS, these nitrobenzylic carbanions can be directly introduced into further reactions with electrophilic reagents: alkylation,⁵ Wittig–Horner reaction,⁶ nitroarylation,⁷ etc.



VNS reaction of α, α -dichlorocarbanions such as those generated by deprotonation of alkyl dichloroacetates^{8a} or dichloromethyl phenyl sulfone^{8b} with nitroarenes gives nitrobenzylic α -chlorocarbanions that, upon protonation, produces typical VNS products.⁸ It was already reported that such nitrobenzylic α -chlorocarbanions, products of VNS reaction between nitroarenes and carbanions of dichloromethyl oxazoline, can enter the Darzens condensation with aromatic aldehydes to give oxiranes.⁹

We have, therefore, expected that nitrobenzylic α -chlorocarbanions produced in the VNS reaction between alkyl dichloroacetates and nitroarenes should be able to add to electron-deficient alkenes—Michael acceptors giving as the ultimate products—esters of cyclopropane carboxylic acids substituted with nitroaromatic rings.

RESULTS AND DISCUSSION

In the preliminary experiments, we have attempted reactions of *t*-butyl α -chloro- α -(4-nitrophenyl)acetate, isolated product of VNS between nitrobenzene 1 and t-butyl dichloroacetate with acrylonitrile and ethyl acrylate under liquid-solid PTC conditions-in the presence of solid K2CO3 in DMF and tetrabutyl ammonium hydrogen sulfate catalyst.¹⁰ The reaction proceeded smoothly to give substituted cyclopropanes in high yields as mixtures of stereoisomers. These results prompted us to perform a one-pot process: formation of the nitrobenzylic α halocarbanions in the VNS reaction of t-butyl dichloroacetate with nitroarenes, followed by addition of the Michael acceptors to the reaction mixtures. The reaction of nitrobenzene with the bulky carbanion of t-butyl dichloroacetate carried out in the presence of t-BuOK in DMF at -5 °C is a fast process, completed within 5 min, and proceeds selectively at the para position. After addition of acrylonitrile or ethyl acrylate to the

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Scheme 1. Vicarious Nucleophilic Substitution in Nitroarenes with α -Chlorocarbanions⁴



Scheme 2. One-Pot Synthesis of Substituted Cyclopropanes via VNS-Michael Addition



Table 1. Results of the One-Pot Reaction of Nitroarenes with Carbanions of *tert*-Butyl Dichloroacetate and Michael Acceptors (Scheme 2)

	Х	Z	Y	R	reaction time [min]	no.	total yield $[\%]^a$	ratio of stereoisomers b
1	CH	Н	CN	Н	60	1a	70	1.3:1
2			COOEt	Н	180	1b	81	6.7:1
3			COOEt	COOEt	360	1c	66	1
4	CH	F	CN	Н	90	2a	59	1.2:1
5			COOEt	Н	180	2b	56	1
6			COOEt	COOEt	480	2c	39	1
7	CH	Cl	CN	Н	90	3a	82	1.1:1
8			COOEt	Н	180	3b	63	1
9			COOEt	COOEt	360	3c	68	1
10	CH	Br	CN	Н	90	4a	82	1.3:1
11			COOEt	Н	180	4b	84	7.6:1
12			COOEt	COOEt	360	4c	85	1
13	CH	OMe	CN	Н	30	5a	71	1:1
14			COOEt	Н	60	5b	78	8.5:1
15			COOEt	COOEt	120	5c	70	1
16	Ν	OEt	CN	Н	60	6a	85	1.9:1
17			COOEt	Н	180	6b	72	1
18			COOEt	COOEt	480	6c	54	1
^a Isolated	yield under	standard con	nditions. ^b Calcu	ilated on the b	asis of ¹ H NMR spectra	and/or m	ass of isolated produc	cts

reaction mixtures, the reaction was continued at room temperature for 2-3 h to give mixtures of stereoisomeric substituted cyclopropanes similar to those obtained under the solid-liquid PTC conditions. According to the identical standard procedure, reactions of *t*-butyl dichloroacetate and acrylonitrile, ethyl acrylate and diethyl maleate with 2-fluoro-, 2-chloro-, 2-bromo-, and 2-methoxynitrobenzenes **2**, **3**, **4** and **5** were carried out. In all of these cases, expected substituted cyclopropanes were obtained in good yields as single stereoisomers or mixtures of stereoisomers (Scheme 2, Table 1).

The reaction with *o*-fluoronitrobenzene 2 carried out under the standard conditions gave somewhat lower yields of the cyclopropanes 2a, 2b and 2c. It appears that this is due to very facile nucleophilic substitution of fluorine located *ortho* to the nitro group; hence, during the reaction, particularly in the second part when the reaction mixtures are kept at room temperatures for a longer time, the side process of S_NAr could proceed.

VNS with *t*-butyl dichloroacetate proceeds also in heteroaromatic compounds that contain nitro groups: nitropyridines, nitrothiophene,^{8a,c} and nitroimidazole;^{8d} hence, we have attempted to synthesize cyclopropanes substituted with nitropyridine moiety. Indeed, the reaction of 2-ethoxy-5-nitropyridine **6** with *t*-butyl dichloroacetate, followed by the reaction of the produced α -chlorocarbanion with acrylonitrile, ethyl acrylate, and diethyl maleate carried out under standard conditions, gave the expected cyclopropanes containing nitropyridyl substituent.

From the results presented in Table 1, it can be seen that reactions of α -halocarbanions generated via VNS of nitroarenes with *t*-butyl dichloroacetate with acrylonitrile to produce cyclopropanes **1a**–**6a** are not stereoselective. Isomers *cis* and *trans* are formed in almost equal quantities. On the other hand, the reaction with ethyl acrylates is highly stereoselective. The ratio of stereoisomers of products **1b**–**6b** is around 8:1 or only one isomer is formed, whereas the reaction with diethyl maleate proceeds stereoselectively, giving single stereoisomers **1c**–**6c**.

The ratio of the stereoisomers can reflect the stereochemistry of the intramolecular 1,3-substitution in the γ -chlorocarbanion—the Michael adducts or can be affected by epimerization of the C–H acidic chiral center of the products in the strongly basic reaction media. When separated major stereoisomer **2a**' was exposed to a solution of *t*-BuOK in DMF, rapid epimerization was observed. We can, therefore, suppose that the ratio of stereoisomers is controlled thermodynamically.

The structure of the products was established on the basis of high-resolution ¹H NMR spectra. Since analysis of values of the chemical shifts and coupling constants was insufficient for determination of the mutual location of the functional groups CN and COOtBu in 1a-6a, and COOEt and COOtBu in 1b-6b, this question was solved using NOE. Thus, irradiation of selected protons of the cyclopropane rings affected protons of the aromatic rings; thus, the mutual location of the functional groups could be unambiguously established. On this basis, the major isomer 3a' was assigned *cis* geometry (CN and COOtBu are on the one side of the cyclopropane ring), whereas 3a'' is a *trans* isomer. Spectra of other isomeric products major isomers 1a'-6a' and minor isomers 1a''-6a'' are very similar to those of 3a' and 3a''; hence, we consider that 1a'-6a' have *cis* and 1a''-6a'' and 1a''-6a'' have *trans* geometry (Scheme 3).





Similar analysis of the coupling constants and NOE effects for 4b' and 4b'' revealed that the major isomer 4b' has *trans* geometry; hence, all major or single isomers 1b'-6b' have *trans* and minor 1b'', 4b'', 5b'' have *cis* geometry (Scheme 4).

Products of the reaction of diethyl maleate 1c-6c are formed as single stereoisomers. On the basis of coupling constants of the cyclopropane ring protons in the ¹H NMR spectra that, for all products, were in the range of 6.1–6.5 Hz, we consider that,

Scheme 4. Stereostructures of the Products 1b–6a Determined by ¹H NOESY Analysis



in all 1c-6c, the ethoxycarbonyl groups are mutually *trans* located (Scheme 5).

Scheme 5. Stereostructures of the Products 1c-6c Determined by ¹H NOESY Analysis



A detailed discussion of the ¹H NMR and NOE effects, that is the basis of structural assignment, is presented in the Supporting Information.

In order to expand the scope of this cyclopropane synthesis and show its general character, we have attempted the reaction of the α -chloronitrobenzylic carbanions generated via VNS reaction with a cyclic Michael acceptor cyclopent-2-en-1-one. The reaction of *o*-bromonitrobenzene 4 and *o*-nitroanisole 5 with *t*-butyl dichloroacetate, followed by addition of cyclopent-2-en-1-one carried out under the standard conditions, gave expected substituted cyclopentanonecyclopropanes in high yields (Scheme 6).

The reported earlier reaction of cyclopent-2-en-1-one with the carbanion of methyl dichloroacetate proceeded with high diastereoselectivity.¹¹ The reaction of this Michael acceptor with α -chloronitrobenzylic carbanions generated via VNS was also diastereoselective.

Geometry of the products **4d** and **5d** was determined on the basis of ¹H NMR spectra using NOE effects. Irradiation of the cyclopropane rings protons δ 2.25 ppm and δ 2.26 ppm results in weak, but still measurable, NOE on the aromatic protons. On this basis, **4d** and **5d** was assigned *cis* geometry.

The bulky tertiary carbanion of *t*-butyl dichloroacetate reacts with nitroarenes selectively at position para; thus, the pnitroaryl substituted cyclopropanes were obtained with high vield in the reaction with nitroarenes that contain substituents in position ortho to the nitro group. Attempts to perform VNS with this bulky carbanion in p-chloronitrobenzene, that can proceed only ortho to the nitro group, gave negative results. On the other hand, the somewhat less bulky carbanion of ethyl dichloroacetate reacts with *p*-chloronitrobenzene 7, giving VNS product in position *ortho* to the nitro group in good yield.^{8a} We have, therefore, attempted to synthesize o-nitroaryl substituted cyclopropane in the reaction of this product with acrylonitrile. The reaction was carried out under typical liquid-solid PTC conditions: K_2CO_3 in DMF and $Bu_4N^+HSO_4^-$ catalyst. Unfortunately, the expected cyclopropane 7a was formed with low yield (8%), and the second product formed also in low yield (13%) was the N-oxide of substituted quinoline 7aa (Scheme 7). It indicates that further reaction of the γ -chloro- α cyano carbanion generated in the Michael addition of the VNS product to acrylonitrile proceeds along two pathways, intramolecular substitution to form the cyclopropane and rapid addition to the nitrogen atom of the vicinal nitro group, followed by elimination of OH⁻ and HCl.

Formation of the quinoline *N*-oxide resembles the reaction of *o*-nitrobenzyl phenyl sulfones with dimethyl maleate and fumarate giving quinoline *N*-oxides in good yield.¹²

Not all mixtures of 1a-6a were separated to give individual stereoisomers. 2a and 6a were separated, so the individual major and minor isomers 2a' and 2a'', 6a' and 6a'' were

Scheme 6. One-Pot Synthesis of Bicyclic Cyclopropanes via VNS-Michael Addition



Scheme 7. Reaction of Ethyl Chloro(2-nitro-5-chlorophenyl)acetate with Acrylonitrile Carried Out under PTC Conditions



obtained. From the mixture of 3a, the individual major stereoisomer 3a' was isolated. The individual products were subjected to spectral and elemental analyses. In the remaining cases, the mixtures were analyzed. From the ¹H NMR spectra of the mixtures, it was possible to subtract spectra of individual major or minor isomers. Products 2b, 3b, and 6b were formed as single stereoisomers, and the samples of major isomers of 1b', 4b', and 5b' were separated from the product mixtures. Thus, all individual major isomers of 1b-6b were obtained and analyzed. The minor isomers 1b", 4b", and 5b" were not isolated. From the spectra of the residual mixtures, after isolation of the major isomers, it was possible to subtract spectra of the minor isomers. All spectral data, scans of NMR spectra of all compounds, and ¹H NMR signals of cyclopropane ring protons in tabular form are given in the experimental section in the Supporting Information.

CONCLUSION

We have shown that *p*-nitrobenzylic α -chlorocarbanions readily produced in the VNS reaction of *t*-butyl dichloroacetate with nitroarenes add to a variety of Michael acceptors to generate γ chlorocarbanions that enter intramolecular substitution, giving esters of cyclopropane carboxylic acids containing *p*-nitroaryl substituents.

This simple generation of the active intermediates opens a versatile way of synthesis of a variety of substituted cyclopropanes and expands the applications of S_NArH reaction in organic synthesis.¹⁴

EXPERIMENTAL SECTION

General Information. The ¹H and ¹³C NMR spectra were recorded at a temperature of 298 K in $CDCl_3$ solutions (in some cases, C_6D_6 was also used) with a 500 MHz apparatus (500 and 125 MHz, for ¹H and ¹³C, respectively). To assign the structures under consideration, the following 1D and 2D experiments were employed: ¹H selective NOESY 2D: COSY and ¹H-¹³C gradient selected HSQC. The ¹H and ¹³C NMR chemical shifts are given relative to the TMS signal at 0.0 ppm. The concentration of solutions used for measurements was about 15–30 mg of compounds in 0.6 mL of solvent. Mass spectra were recorded using an electron spray ionization (ESI) technique. Melting point temperatures were determinated with a

heating rate of 5 °C/min and were uncorrected. Column chromatography was performed using silica gel 60 (0.040–0.063 mm). Thin-layer chromatography was performed on precoated silica gel plates and visualized under a UV lamp. For column chromatography, hexane/ethyl acetate mixtures were used as the eluents. Solid cyclopropanes were recrystallized from hexane/Et₂O. All solvents were distilled before use. Dry DMF was obtained by distillation from CaH₂. *tert*-Butyl dichloroacetate, ethyl chloro(2-nitro-S-chlorophenyl)acetate, and *t*-butyl chloro(4-nitrophenyl)acetate were prepared according to a literature procedure adequately.^{8a,c} All other chemicals were commercial and used as received.

General Procedure for Synthesis of Substituted Cyclopropanes. To a suspension of potassium t-butoxide (243 mg, 2.1 mmol) in dry DMF (3 mL) cooled to -5 °C under argon, a solution of a nitroarene (1.0 mmol) and tert-butyl dichloroacetate (206 mg, 1.1 mmol) in DMF (1 mL) was added at once. The mixture was stirred at -5 °C for 5 min, and DMF (5 mL) and a solution of a Michael acceptor (1.5 mmol) in DMF (2 mL) were added. The cooling bath was removed, and the mixture was stirred at room temperature until VNS product was consumed (TLC control). The reaction mixture was poured in water (20 mL), and the product was extracted with dichloromethane $(3 \times 15 \text{ mL})$. The combined extract was washed with water $(3 \times 20 \text{ mL})$ and dried, and the solvent was evaporated. The products were purified by column chromatography on silica gel, hexane-ethyl acetate eluent. Evaporation of the solvent gave final products as single or a mixture of stereoisomers. Ratio of stereoisomers was determined by ¹H NMR using integration of diagnostic signals of methylene protons.

1a. Yield 203 mg, 70%, ratio of stereoisomers = 1.3:1, not separated, solidified upon standing. From ¹H NMR spectra of the mixture, signals of both isomers were assigned.

1a'. ¹H NMR (500 Hz, CDCl₃) δ (ppm) 1.47 (s, 9H, *t*Bu), 1.71 (dd, J = 9.4 Hz, J = 5.4 Hz, 1H, CH₂), 2.06–2.08 (m, 1H, CHCN), 2.30 (dd, 1H, J = 6.3 Hz, J = 5.4 Hz, CH₂), 7.53 (d, J = 8.7 Hz, 2H, ArH), 8.21 (d, J = 8.7 Hz, 2H, ArH).

1*a*". ¹H NMR (500 Hz, CDCl₃) δ (ppm) 1.37 (s, 9H, tBu), 1.82– 1.84 (m, 1H, CH₂), 2.04–2.06 (m, 1H, CH₂), 2.53 (dd, J = 9.4 Hz, J = 6.4 Hz, 1H, CHCN), 7.56 (d, J = 8.7 Hz, 2H, ArH), 8.26 (d, J = 8.7 Hz, 2H, ArH).

Analysis of the Mixtures of Isomers 1a' and 1a". 13 C NMR (125 Hz, CDCl₃) δ 12.7, 13.0, 19.5, 20.4, 27.7, 27.8, 35.8, 36.8, 83.8, 84.3, 117.1, 117.4, 123.7, 123.7, 130.9, 131.7, 140.9, 143.5, 147.7, 166.6, 168.2; Anal. Calcd for C₁₅H₁₆N₂O₄: C, 62.49; H, 5.59; N, 9.72. Found: C, 62.46; H, 5.70; N, 9.70; MS (ESI, MeOH): 311 [M + Na]⁺, 599 [2M + Na]⁺.

1b. Yield 271 mg, 81%, ratio of stereoisomers = 6.7:1. Major isomer **1b**' isolated by double recrystallization.

1b'. White solid, mp = 81.5–82.5 °C; FT-IR (KBr, ν_{max}/cm^{-1}) 2981, 1734, 1717, 1601, 1515, 1474, 1455, 1392, 1382, 1372, 1366, 1340, 1294, 1280, 1258, 1203, 1159, 1105, 1028, 985, 854, 843, 803, 752, 699, 533; ¹H NMR (500 Hz, CDCl₃) δ (ppm) 1.04 (t, *J* = 7.1 Hz, 3H, <u>CH₃-CH₂</u>), 1.37 (s, 9H, tBu), 1.91 (dd, *J* = 8.5 Hz, *J* = 4.5 Hz, 1H, CH₂), 1.98, (dd, *J* = 6.4 Hz, *J* = 4.5 Hz, 1H, CH₂), 2.72 (dd, *J* = 8.5 Hz, *J* = 6.5 Hz, 1H, <u>CH</u>-COOEt), 3.88–3.95 (m, 2H, CH₃-<u>CH₂</u>), 7.42 (d, *J* = 8.7 Hz, 2H, ArH), 8.16 (2H, d, *J* = 8.7 Hz, ArH); ¹³C NMR (125 Hz, CDCl₃) δ 14.0, 19.1, 27.8, 29.20, 37.4, 61.1, 82.6, 123.0, 131.5, 143.0, 147.2, 168.8, 169.7; Anal. Calcd for C₁₇H₂₁NO₆: C, 60.89; H, 6.31; N, 4.18. Found: C, 60.98; H, 6.34; N, 4.05; MS (ESI, MeOH): 358 [M + Na]⁺, 693 [2M + Na]⁺.

1b". NMR spectra were subtracted from the spectra of the mixtures. ¹H NMR (500 Hz, CDCl₃) δ (ppm) 1.32 (t, *J* = 7.1 Hz, 3H, <u>CH₃</u>-CH₂), 1.38 (s, 9H, tBu), 1.49 (dd, *J* = 8.5 Hz, *J* = 5.1 Hz, 1H, CH₂), 2.15–2.18 (m, 1H, CH₂), 2.23, (dd, *J* = 8.5 Hz, *J* = 6.6 Hz, 1H, <u>CH</u>-COOEt), 4.20–4.25 (m, 2H, CH₃-<u>CH₂</u>), 7.58 (d, *J* = 8.8 Hz, 2H, ArH), 8.18 (d, *J* = 8.8 Hz, 2H, ArH); ¹³C NMR (125 Hz, CDCl₃) δ 14.3, 18.7, 27.7, 28.5, 39.0, 61.3, 82.3, 123.6, 130.0, 142.8, 146.1, 167.4, 169.6.

1c. Yield 271 mg, 66%, pale yellow oil; FT-IR (film, ν_{max}/cm^{-1}) 2980, 1723, 1605, 1521, 1446, 1393, 1369, 1346, 1269, 1257, 1208, 1146, 1113, 1095, 1080, 1035, 1014, 904, 853, 847, 789, 749, 737, 695, 668, 654; ¹H NMR (500 Hz, CDCl₃) δ (ppm) 1.09 (t, *J* = 7.2 Hz, 3H, <u>CH₃-CH₂</u>), 1.31–1.34 (m, 12H, 3H from <u>CH₃CH₂</u> and 9H from tBu), 3.00 (d, *J* = 6.2 Hz, 1H, <u>CH</u>-COOEt), 3.22 (d, *J* = 6.2 Hz, 1H, <u>CH</u>-COOEt), 3.95–3.99 (m, 2H, CH₃-<u>CH₂</u>), 4.22–4.26 (m, 2H, CH₃-<u>CH₂</u>), 7.49–7.51 (m, 2H, ArH), 8.16–8.18 (m, 2H, ArH); ¹³C NMR (125 Hz, CDCl₃) δ 14.0, 14.2, 27.6, 31.8, 32.0, 44.9, 61.5, 61.7, 83.1, 123.6, 130.4, 141.9, 147.6, 165.9, 167.5, 167.7; Anal. Calcd for C₂₀H₂₅NO₈: C, 58.96; H, 6.18; N, 3.44. Found: C, 58.87; H, 6.01; N, 3.36; MS (ESI, MeOH): 430 [M + Na]⁺, 837 [2M + Na]⁺.

2a. Yield 182 mg, 59%, ratio of stereoisomers = 1.2:1, separated by column chromatography.

2a'. White solid, mp = 120.5–122.5 °C; FT-IR (KBr, ν_{max}/cm^{-1}) 3118, 2988, 2243, 1722, 1610, 1602, 1526, 1489, 1457, 1434, 1418, 1393, 1370, 1345, 1313, 1258, 1220, 1203, 1155, 1109, 1084, 954, 938, 888, 864, 843, 830, 792, 750, 736, 693, 672, 626, 552, 455; ¹H NMR (500 Hz, CDCl₃) δ (ppm) 1.48 (s, 9H, tBu), 1.72 (dd, J = 9.5 Hz, J = 5.5 Hz, 1H, CH₂), 2.06 (dd, J = 9.5 Hz, J = 6.5 Hz, 1H, CH–CN), 2.31 (dd, J = 6.5, J = 5.5, 1H, CH₂), 7.29–7.32 (m, 2H, ArH), 8.04–8.07 (m, 1H, ArH); ¹³C NMR (125 Hz, CDCl₃) δ 12.9, 19.6, 27.8, 36.2 (d, ⁴ $J_{CF} = 1.2$ Hz), 84.7, 116.7, 120.1 (d, ² $J_{CF} = 21.2$ Hz), 126.0 (d, ³ $J_{CF} = 4.0$ Hz), 126.3 (d, ⁴ $J_{CF} = 2.4$ Hz), 136.9 (d, ³ $J_{CF} = 6.9$ Hz), 144.9 (d, ² $J_{CF} = 8.1$ Hz), 155.1 (d, ¹ $J_{CF} = 265.5$ Hz), 166.1; Anal. Calcd for C₁₅H₁₅FN₂O₄: C, 58.82; H, 4.92; F, 6.20; N, 9.15; Found: 58.80; H, 4.91; F, 6.23; N, 9.09; MS (ESI, MeOH): 329 [M + Na]⁺, 635 [2M + Na]⁺.

2a". White solid, mp = 112.5–114.5 °C; FT-IR (KBr, ν_{max}/cm^{-1}) 3057, 2982, 2248, 1721, 1612, 1599, 1530, 1494, 1429, 1415, 1371, 1356, 1337, 1307, 1286, 1226, 1158, 1106, 1085, 939, 889, 839, 775, 750, 734, 727, 702, 636, 560, 462; ¹H NMR (500 Hz, CDCl₃) δ (ppm) 1.38 (s, 9H, tBu), 1.81 (dd, *J* = 6.3 Hz, *J* = 5.1 Hz, 1H, CH₂), 2.06 (dd, *J* = 9.4 Hz, *J* = 5.1 Hz, 1H, CH₂), 2.53 (dd, *J* = 9.4, *J* = 6.3, 1H, CH-CN), 7.26–7.33 (m, 2H, ArH), 8.09–8.12 (m, 1H, ArH); ¹³C NMR (125 Hz, CDCl₃) δ 13.2, 20.4, 27.7, 35.5 (d, ⁴*J*_{CF} = 1.2 Hz), 84.2, 117.1, 120.8 (d, ²*J*_{CF} = 21.2 Hz), 126.3 (d, ⁴*J*_{CF} = 2.5 Hz), 126.8 (d, ³*J*_{CF} = 5.0 Hz), 137.1 (d, ³*J*_{CF} = 7.5 Hz), 142.4 (d, ²*J*_{CF} = 8.8 Hz), 155.2 (d, ¹*J*_{CF} = 263.8 Hz), 167.6; Anal. Calcd for C₁₅H₁₅FN₂O₄: C, 58.82; H, 4.92; F, 6.20; N, 9.15. Found: C, 58.62; H, 4.94; F, 6.13; N, 9.09; MS (ESI, MeOH): 329 [M + Na]⁺, 635 [2M + Na]⁺.

2b'. Yield 197 mg, 56%, white solid, mp = 76–77 °C; FT-IR (KBr, ν_{max}/cm^{-1}) 2980, 2940, 1734, 1721, 1612, 1602, 1520, 1491, 1455, 1384, 1372, 1366, 1342, 1281, 1252, 1229, 1281, 1196, 1157, 1109, 1083, 1030, 991, 949, 842, 831, 761, 753, 693, 673, 593, 560; ¹H NMR (500 Hz, CDCl₃) δ (ppm) 1.10 (t, *J* = 7.1 Hz, 3H, <u>CH₃-CH₂</u>), 1.38 (s, 9H, tBu), 1.91 (dd, *J* = 8.5 Hz, *J* = 4.6 Hz, 1H, CH₂), 1.94 (dd, *J* = 6.5 Hz, *J* = 4.6 Hz, 1H, CH₂), 2.71 (dd, *J* = 8.5 Hz, *J* = 6.5 Hz, 1H, <u>CH₂</u>)

COOEt), 3.99–3.94 (m, 2H, CH₃-<u>CH₂</u>), 7.26–7.17 (m, 2H, ArH), 8.01–7.98 (m, 1H, ArH); ¹³C NMR (125 Hz, CDCl₃) δ 14.0, 19.2, 27.8, 29.3, 37.1 (d, ⁴J_{CF} = 1.2 Hz), 61.3, 83.0, 120.6 (d, ²J_{CF} = 21.2 Hz), 125.5 (d, ⁴J_{CF} = 2.5 Hz), 126.8 (d, ³J_{CF} = 3.7 Hz), 136.3 (d, ³J_{CF} = 6.2 Hz), 144.7 (d, ²J_{CF} = 8.8 Hz), 155.0 (d, ¹J_{CF} = 262.5 Hz), 168.7, 169.1; Anal. Calcd for C₁₇H₂₀FNO₆: C, 57.79; H, 5.71; F, 5.38; N, 3.96. Found: C, 57.80; H, 5.74; F, 5.39; N, 3.92. MS (ESI, MeOH): 376 [M + Na]⁺, 729 [2M + Na]⁺.

2c. Yield 165 mg, 39%, pale yellow oil; FT-IR (film, ν_{max}/cm^{-1}) 2981, 1724, 1613, 1601, 1528, 1491, 1447, 1418, 1394, 1369, 1346, 1275, 1257, 1197, 1143, 1097, 1035, 976, 895, 861, 835, 787, 751, 730, 688, 668, 655; ¹H NMR (500 Hz, CDCl₃) δ (ppm) 1.14 (t, J = 7.1 Hz, 3H, <u>CH₃-CH₂</u>), 1.32–1.36 (m, 12H, 3H from <u>CH₃-CH₂</u> and 9H from tBu), 2.97 (d, J = 6.2 Hz, 1H, <u>CH</u>-COOEt), 3.21 (d, J = 6.2 Hz, 1H, <u>CH</u>-COOEt), 4.02 (q, J = 7.2 Hz, 2H, CH₃-<u>CH₂</u>), 4.24 (q, J = 7.1, 2H, CH₃-<u>CH₂</u>), 7.25–7.29 (m, 2H, ArH), 8.01–8.04 (m, 1H, ArH); ¹³C NMR (125 Hz, CDCl₃) δ 14.0, 14.2, 27.6, 31.9, 32.2, 44.5 (d, ⁴ $J_{CF} = 1.2$ Hz), 61.7, 61.9, 83.6, 119.6 (d, ² $J_{CF} = 21.2$ Hz), 125.6 (d, ³ $J_{CF} = 3.8$ Hz), 126.1 (d, ⁴ $J_{CF} = 2.5$ Hz), 136.8 (d, ³ $J_{CF} = 7.5$ Hz), 143.5 (d, ² $J_{CF} = 8.8$ Hz), 155.2 (d, ¹ $J_{CF} = 263.8$ Hz), 165.5, 167.4, 167.4; Anal. Calcd for C₂₀H₂₄FNO₈: C, 56.47; H, 5.69; F, 4.47; N, 3.29. Found: C, 56.47; H, 5.77; F, 4.42; N, 3.30; MS (ESI, MeOH): 448 [M + Na]⁺, 873 [2M + Na]⁺.

3a. Yield 265 mg, 82%, ratio of stereoisomers = 1.1:1. Major isomer 3a' isolated by column chromatography.

3a'. White solid, mp = 118–120 °C; FT-IR (KBr, $\nu_{\rm max}/\rm cm^{-1}$) 3103, 2988, 2247, 1719, 1593, 1580, 1524, 1475, 1436, 1371, 1364, 1354, 1320, 1287, 1255, 1214, 1155, 1141, 1102, 1051, 930, 919, 864, 848, 837, 815, 805, 769, 741, 700, 663, 623, 566, 482; ¹H NMR (500 Hz, CDCl₃) δ (ppm) 1.48 (s, 9H, tBu), 1.71 (dd, *J* = 9.4 Hz, *J* = 5.3 Hz, 1H, CH₂), 2.04 (dd, *J* = 9.4 Hz, *J* = 6.8 Hz, 1H, CH-CN), 2.29–2.31 (m, 1H, CH₂), 7.41 (dd, *J* = 8.4 Hz, 1H, ArH); ¹³C NMR (125 Hz, CDCl₃) δ 12.8, 19.4, 27.8, 36.1, 84.7, 116.8, 125.7, 127.3, 129.2, 133.3, 142.4, 147.4, 166.2; Anal. Calcd for C₁₅H₁₅ClN₂O₄: C, 55.82; H, 4.68; Cl, 10.98; N, 8.68; Found: C, 55.75; H, 4.71; Cl, 10.82; N, 8.51; MS (ESI, MeOH): 345 [M + Na]⁺.

3*a*". NMR spectra were subtracted from the spectra of the mixtures. ¹H NMR (500 Hz, CDCl₃) δ (ppm) 1.38 (s, 9H, *t*Bu), 1.82 (dd, *J* = 6.4 Hz, *J* = 5.2 Hz, 1H, CH₂), 2.06 (dd, *J* = 9.4 Hz, *J* = 5.2 Hz, 1H, CH₂), 2.53 (dd, *J* = 9.4, *J* = 6.4, 1H, CH-CN), 7.44 (dd, *J* = 8.4 Hz, *J* = 1.8 Hz 1H, ArH), 7.56 (d, *J* = 1.8 Hz, 1H, ArH), 7.93 (d, *J* = 8.4 Hz, 1H, ArH); ¹³C NMR (125 Hz, CDCl₃) δ 13.1, 20.3, 27.7, 35.3, 84.2, 117.1, 125.8, 127.3, 130.0, 134.0, 139.9, 147.5, 167.7.

3b'. Yield 235 mg, 63%, pale yellow oil; FT-IR (film, ν_{max}/cm^{-1}) 2979, 1716, 1595, 1582, 1527, 1478, 1457, 1393, 1381, 1368,1346, 1274, 1253, 1200, 1151, 1093, 1049, 1033, 986, 920, 861, 839, 802, 749, 738, 699, 668, 654; ¹H NMR (500 Hz, CDCl₃) δ (ppm) 1.09 (t, *J* = 7.1 Hz, 3H, <u>CH₃-CH₂</u>), 1.39 (s, 9H, tBu), 1.91 (dd, *J* = 8.5 Hz, *J* = 4.6 Hz, 1H, CH₂), 1.95 (dd, *J* = 6.5 Hz, *J* = 4.6 Hz, 1H, CH₂), 2.70 (dd, *J* = 8.5 Hz, *J* = 6.5 Hz, *I* = 1.8, 1H, ArH), 7.44 (d, *J* = 1.8 Hz, 1H, ArH), 7.84 (d, *J* = 8.4 Hz,1H, ArH); ¹³C NMR (125 Hz, CDCl₃) δ 14.0, 19.0, 27.8, 29.2, 36.9, 61.2, 82.9, 125.2, 126.7, 129.8, 134.0, 142.0, 146.7, 168.6, 169.2; Anal. Calcd for C₁₇H₂₀ClNO₆: C, 55.21; H, 5.45; Cl, 9.59; N, 3.79, Found: C, 55.11; H, 5.35; Cl, 9.73; N, 3.83; MS (ESI, MeOH): 392 [M + Na]⁺, 761 [2M + Na]⁺.

3c. Yield 301 mg, 68%, pale yellow oil; FT-IR (film, ν_{max}/cm^{-1}) 2981, 1723, 1595, 1581, 1530, 1446, 1393, 1369, 1338, 1272, 1254, 1209, 1146, 1094, 1035, 864, 839, 803, 787, 749, 668; ¹H NMR (500 Hz, CDCl₃) δ (ppm) 1.13 (t, J = 7.1 Hz, 3H, <u>CH₃-CH₂</u>), 1.32–1.36 (m, 12H, 3H from <u>CH₃CH₂</u> and 9H from tBu), 2.97 (d, J = 6.2 Hz, 1H, <u>CH</u>-COOEt), 3.21 (d, J = 6.2 Hz, 1H, <u>CH</u>-COOEt), 4.00–4,05 (m, 2H, CH₃-<u>CH₂</u>), 4.24 (q, J = 7.1 Hz, 2H, CH₃-<u>CH₂</u>), 7.37 (dd, J = 8.5 Hz, J = 2.0 Hz, 1H, ArH), 7.51 (d, J = 1.8 Hz, 1H, ArH), 7.84 (d, J = 8.4 Hz 1H, ArH); ¹³C NMR (125 Hz, CDCl₃) δ 14.0, 14.2, 27.6, 31.8, 32.1, 44.3, 61.7, 61.8, 83.5, 125.7, 127.2, 128.7, 132.9, 140.9, 147.2, 165.6, 167.4, 167.5; Anal. Calcd for C₂₀H₂₄ClNO₈: C, 54.36; H, 5.47; Cl, 8.02; N, 3.17. Found: C, 54.10; H, 5.52; Cl, 8.05; N, 3.12; MS (ESI, MeOH): 464 [M + Na]⁺, 905 [2M + Na]⁺.

4a. Yield 302 mg, 82%, ratio of stereoisomers = 1.3:1, not separated, solidified upon standing.

From ¹H NMR spectra of the mixture, signals of both isomers were assigned:

4a'. ¹H NMR (500 Hz, CDCl₃) δ (ppm) 1.48 (s, 9H, *t*Bu), 1.71 (dd, J = 9.4 Hz, J = 5.4 Hz, 1H, CH₂), 2.03 (dd, J = 9.4 Hz, J = 6.7 Hz, 1H, CH-CN), 2.29 (1H, dd, J = 6.7 Hz, J = 5.4 Hz, CH₂), 7.45 (dd, J = 8.4 Hz, J = 1.7 Hz, 1H, ArH), 7.72 (d, J = 1.7 Hz, 1H, ArH), 7.83 (d, J = 8.4 Hz, 1H, Ar-H).

4*a*". ¹H NMR (500 Hz, CDCl₃) δ (ppm) 1.38 (s, 9H, tBu), 1.80– 1.83 (m, 1H, CH₂), 2.06 (dd, J = 9.4 Hz, J = 5.1 Hz, 1H, CH₂) 2.51 (dd, J = 9.4, J = 6.4, 1H, CH-CN), 7.48 (dd, J = 8.4 Hz, J = 1.7 Hz, 1H, ArH), 7.75 (d, J = 1.7 Hz, 1H, ArH), 7.90 (d, J = 8.4 Hz, 1H, ArH).

Analysis of the Mixtures of Isomers **4a**' and **4a**". ¹³C NMR (125 Hz, CDCl₃) δ 12.8, 13.1, 19.4, 20.3, 27.7, 27.8, 35.2, 36.0, 84.1, 84.7, 114.6, 114.6, 116.8, 117.1, 125.6, 125.7, 129.92, 130.7, 136.5, 137.2, 139.8, 142.3, 149.3, 166.2, 167.7; Anal. Calcd for C₁₅H₁₅BrN₂O₄: C, 49.06; H, 4.12; Br, 21.76; N, 7.63. Found: C, 49.26; H, 4.08; Br, 21.99; N, 7.43; MS (ESI, MeOH): 389 [M + Na]⁺.

4b. Yield 350 mg, 84%, ratio of stereoisomers = 7.6:1. Major isomer **4b**' isolated by column chromatography; the residue was a 1.1:1 mixture of isomers.

4b'. Pale yellow oil; FT-IR (film, ν_{max}/cm^{-1}) 2979, 1716, 1594, 1578, 1528, 1476, 1457, 1394, 1380, 1368, 1354, 1199, 1149, 1093, 1038, 985, 917, 858, 839, 799, 749, 735, 698, 668, 652; ¹H NMR (500 Hz, CDCl₃) δ (ppm) 1.08 (t, J = 7.1 Hz, 3H, <u>CH₃-CH₂</u>), 1.38 (s, 9H,tBu), 1.90 (dd, J = 8.5 Hz, J = 4.6 Hz, 1H, CH₂), 1.96 (dd, J = 6.4 Hz, J = 4.6 Hz, 1H, CH₂), 2.69 (dd, J = 8.5 Hz, J = 6.4 Hz, 1H, <u>CH-COOEt</u>), 3.91- 4.00 (m, 2H, CH₃-<u>CH₂</u>), 7.33 (dd, J = 8.4 Hz, J = 1.8, 1H, ArH), 7.63 (1H, d, J = 1.7 Hz, ArH), 7.81 (1H, d, J = 8.4 Hz, ArH); ¹³C NMR (125 Hz, CDCl₃) δ 14.1, 19.0, 27.8, 29.2, 36.8, 61.26, 82.9, 114.0, 125.2, 130.5, 137.2, 141.9, 148.5, 168.6, 169.2; Anal. Calcd for C₁₇H₂₀BrNO₆: C, 49.29; H, 4.87; Br, 19.29; N, 3.38; Found: C, 49.11; H, 4.83; Br, 19.23; N, 3.45; MS (ESI, MeOH): 436 [M + Na]⁺, [2M + Na]⁺.

4b". NMR spectra were subtracted from the spectra of the mixtures. ¹H NMR (500 Hz, CDCl₃) δ (ppm) 1.32 (t, *J* = 7.1 Hz, 3H, <u>CH₃</u>-CH₂), 1.39 (s, 9H, tBu), 1.48 (dd, *J* = 8.6 Hz, *J* = 5.1 Hz, 1H, CH₂), 2.16–2.13 (m, 1H, CH₂), 2.20 (dd, *J* = 8.5 Hz, *J* = 6.7 Hz, 1H, <u>CH</u>-COOEt, 4.18–4.25 (m, 2H, CH₃-<u>CH₂</u>), 7.50 (dd, *J* = 8.4 Hz, *J* = 1.8, 1H, ArH), 7.78 (d, *J* = 1.7 Hz, 1H, ArH), 7.80 (d, *J* = 8.4 Hz, 1H, ArH); ¹³C NMR (125 Hz, CDCl₃) δ 14.2, 18.5, 27.8, 28.6, 38.4, 61.4, 82.6, 114.5, 125.6, 129.0, 135.7, 144.9, 148.9, 167.0, 169.3.

4c. Yield 416 mg, 85%, yellow oil; FT-IR (film, ν_{max}/cm^{-1}) 2981, 1723, 1592, 1577, 1531, 1457, 1393, 1368, 1337, 1271, 1255, 1209, 1145, 1094, 1037, 859, 840, 783, 748, 700, 668; ¹H NMR (500 Hz, CDCl₃) δ (ppm) 1.14 (t, *J* = 7.1 Hz, 3H, <u>CH₃-CH₂</u>), 1.32–1.36 (m, 12H, 3H from <u>CH₃CH₂</u> and 9H from tBu), 2.97 (d, *J* = 6.3 Hz, 1H, <u>CH-COOEt</u>), 3.20 (d, *J* = 6.3 Hz, 1H, <u>CH-COOEt</u>), 4.00–4,05 (m, 2H, CH₃-<u>CH₂</u>), 4.24 (q, *J* = 7.1, 2H, CH₃-<u>CH₂</u>), 7.41 (dd, *J* = 8.4 Hz, *J* = 1.8 Hz, 1H, ArH); ¹³C NMR (125 Hz, CDCl₃) δ 14.1, 14.2, 27.6, 31.8, 32.0, 44.2, 61.7, 61.8, 83.5, 114.5, 125.6, 129.4, 136.0, 140.8, 149.1, 165.6, 167.3, 167.5; Anal. Calcd for C₂₀H₂₄BrNO₈: C, 49.40; H, 4.97; Br, 16.43; N, 2.88; Found: C, 49.18; H, 5.09; Br, 16.34; N, 2.84; MS (ESI, MeOH): 508 [M + Na]⁺, 995 [2M + Na]⁺.

4d. Yield 299 mg, 76%, white solid, mp = 112–115 °C; FT-IR (KBr, ν_{max} /cm⁻¹) 3097, 2972, 1723, 1594, 1576, 1543, 1369, 1323, 1290, 1249, 1161, 1145, 1073, 949, 897, 861, 852, 839, 808, 733, 658, 614, 572, 500, 474; ¹H NMR (500 Hz, CDCl₃) δ (ppm) 0.89–0.97 (m, 1H), 1.35 (s, 9H), 1.93–2.03 (m, 2H), 2.36–2.44 (m, 1H), 2.80–2.87 (m, 2H), 7.36 (d, *J* = 7.6 Hz, 1H), 7.69 (s, 1H), 7.84 (d, *J* = 8.3 Hz, 1H); ¹H NMR (500 Hz, C₆D₆) δ (ppm) 0.37–0.33 (m, 1H, CH₂(C=O)), 1.09–1.04 (m, 10H, 9H from tBu and 1H from CH₂), 1.39–1.33 (m, 2H, 1H from CH₂ and 1H from CH₂(C=O)), 2.25 (t, *J* = 5.7, 1H, CH), 2.66 (d, *J* = 6.1 Hz, 2H, CH(C=O)), 6.70 (d, *J* = 6.0 Hz, 1H, Ar), 7.00 (d, *J* = 6.4 Hz, 1H, ArH), 7.36 (s, 1H, ArH); ¹³C NMR (125 Hz, CDCl₃) δ 20.6, 27.8, 34,8, 35.0, 39.2, 41.6, 83.2, 114.6, 125.8, 130.4, 137.1, 139.8, 149.0, 168.2, 211.5; ¹³C NMR (125 Hz, C₆D₆) δ 20.3, 27.5, 34.5, 35.0, 39.0, 41.5, 82.2, 125.62, 125.65, 130.5,

137.0, 139.7, 149.5, 168.2, 209.5; Anal. Calcd for $C_{17}H_{18}BrNO_5$: C, 51.53; H, 4.58; N, 3.53; Br, 20.17; Found: C, 51.69; H, 4.52; N, 3.53; Br, 20.08; MS (ESI, MeOH): 345 $[M + Na]^+$.

5a. Yield 226 mg, 71%, ratio of stereoisomers = 1:1, not separated. From ¹H NMR spectra of the mixture, signals of both isomers were assigned.

 $\mathbf{5a'}$. ¹H NMR (500 Hz, CDCl₃) δ (ppm) 1.47 (s, 9H, *t*Bu), 1.70 (dd, J = 9.4 Hz, J = 5.3 Hz, 1H, CH₂), 2.02–2.04 (m, 1H, CHCN), 2.29 (dd, 1H, J = 6.6 Hz, J = 5.3 Hz CH₂), 3.99 (s, 3H, OCH₃), 7.04 (dd, 1H, J = 8.3 Hz, J = 1.7 Hz ArH), 7.10 (d, 1H, J = 1.6 Hz, ArH), 7.87 (d, 1H, J = 8.4 Hz, ArH).

5aⁿ. ¹H NMR (500 Hz, CDCl₃) δ (ppm) 1.38 (s, 9H, tBu), 1.79 (dd, J = 6.4 Hz, J = 5.1 Hz, 1H, CH₂), 2.04–2.05 (m, 1H, CH₂), 2.50 (dd, J = 9.4, J = 6.4, 1H, CH-CN), 3.97 (s, 3H, OCH₃), 6.99 (dd, J = 8.3 Hz, J = 1.6 Hz, 1H, ArH), 7.08 (d, J = 1.5 Hz, 1H, ArH), 7.80 (d, J = 8.3 Hz, 1H, ArH).

Analysis of the Mixtures of Isomers **5a**' and **5a**". ¹³C NMR (125 Hz, CDCl₃) δ 12.7, 12.9, 19.5, 20.6, 27.8, 27.9, 36.1, 36.9, 56.7, 83.8, 84.3, 115.3, 116.0, 117.2, 117.5, 121.5, 122.5, 125.9, 126.0, 139.3, 139.4, 140.6, 143.1, 152.8, 152.9, 166.6, 168.1; Anal. Calcd for C₁₆H₁₈N₂O₅: C, 60.37; H, 5.70; N, 8.80; Found: C, 60.50; H, 5.56; N, 8.75; MS (ESI, MeOH): 341 [M + Na]⁺, 659 [2M + Na]⁺.

5b. Yield 284 mg, 78%, ratio of stereoisomers = 8.8:1. Major isomer **5b**' isolated by column chromatography; the residue was a 1.1:1 mixture of isomers.

5b'. Pale yellow oil; FT-IR (film, ν_{max}/cm^{-1}) 2979, 1716, 1607, 1591, 1519, 1492, 1458, 1406, 1381, 1368, 1348, 1303, 1277, 1244, 1197, 1146, 1108, 1085, 1024, 992, 936, 856, 840, 799, 755, 727, 720, 704, 675, 668, 650; ¹H NMR (500 Hz, CDCl₃) δ (ppm) 1.05 (t, *J* = 7.1 Hz, 3H, <u>CH₃-CH₂</u>), 1.38 (s, 9H, tBu), 1.88 (dd, *J* = 8.5 Hz, *J* = 4.6 Hz, 1H, CH₂), 1.96 (dd, *J* = 6.4 Hz, *J* = 4.6 Hz, 1H, CH₂), 2.70 (dd, *J* = 8.4 Hz, *J* = 6.4 Hz, 1H, <u>CH</u>-COOEt), 3.91–3.96 (m, 5H), 6.92 (dd, *J* = 8.4 Hz, *J* = 1.6, 1H, ArH), 6.97 (d, *J* = 1.6 Hz, 1H, ArH), 7.80 (d, *J* = 8.4 Hz, 1H, ArH); ¹³C NMR (125 Hz, CDCl₃) δ 14.0, 19.1, 27.8, 29.2, 37.6, 56.5, 61.1, 82.6, 115.9, 122.6, 125.4, 138.6, 142.7, 152.7, 168.8, 169.6; Anal. Calcd for C₁₈H₂₃NO₇: C, 59.17; H, 6.34; N, 3.83; Found: C, 58.96; H, 6.14; N, 4.04; MS (ESI, MeOH): 388 [M + Na]⁺, 753 [2M + Na]⁺.

5b". NMR spectra were subtracted from the spectra of the mixtures. ¹H NMR (500 Hz, CDCl₃) δ (ppm) 1.32 (t, *J* = 7.1 Hz, 3H, <u>CH₃</u>-CH₂), 1.39 (s, 9H, tBu), 1.48 (dd, *J* = 8.6 Hz, *J* = 5.1 Hz, 1H, CH₂), 2.12 (dd, *J* = 6.5 Hz, *J* = 5.1 Hz, 1H, CH₂), 2.21 (dd, *J* = 8.6 Hz, *J* = 6.5 Hz, 1H, <u>CH</u>-COOEt), 3.97 (s, 3H, CH₃O), 4.18–4.25 (m, 2H, CH₃-CH₂), 7.05 (dd, *J* = 8.4 Hz, *J* = 1.8, 1H, ArH), 7.15 (d, *J* = 1.8 Hz, 1H, ArH), 7.79 (d, *J* = 8.4 Hz, 1H, ArH); ¹³C NMR (125 Hz, CDCl₃) δ 14.2, 18.7, 27.8, 30.9, 39.6, 56.6, 61.2, 82.2, 114.4, 120.7, 125.8, 138.9, 145.6, 152.9, 167.4, 169,8.

5c. Yield 308 mg, 70%, yellow oil; FT-IR (film, ν_{max}/cm^{-1}) 2980, 1724,1607, 1590, 1522, 1495, 1463, 1407,1393, 1368, 1244, 1204, 1185, 1144, 1097, 1029, 908, 876, 859, 834, 785, 755, 728, 668; ¹H NMR (500 Hz, CDCl₃) δ (ppm) 1.10 (t, *J* = 7.1 Hz, 3H, <u>CH₃-CH₂</u>), 1.32–1.36 (m, 12H, 3H from <u>CH₃CH₂</u> and 9H from tBu), 3.00 (d, *J* = 6.1 Hz, 1H, <u>CH</u>-COOEt), 3.20 (d, *J* = 6.1 Hz, 1H, <u>CH</u>-COOEt), 3.95–4,02 (m, 5H), 4.24 (q, *J* = 7.1, 2H, CH₃-<u>CH₂</u>), 7.00–7.02 (m, 2H, ArH), 7.81 (d, *J* = 8.3 Hz, 1H, ArH); ¹³C NMR (125 Hz, CDCl₃) δ 14.0, 14.2, 27.6, 31.7, 31.9, 45.4, 56.7, 61.5, 61.7, 83.1, 114.5, 121.3, 125.8, 139.2, 141.5, 152.9, 165.9, 167.4, 167.9; Anal. Calcd for C₂₁H₂₇NO₉: C, 57.66; H, 6.22; N, 3.20; Found: C, 57.68; H, 5.99; N, 3.08; MS (ESI, MeOH): 460 [M + Na]⁺, 897 [2M + Na]⁺.

5d. Yield 242 mg, 70%, white solid, mp = 146–148 °C; FT-IR (KBr, ν_{max}/cm^{-1}) 2977, 2941, 1719, 1607, 1591, 1514, 1491, 1464, 1406, 1370, 1352, 1302, 1290, 1267, 1254, 1237, 1162, 1146, 1095, 1076, 1056, 1028, 944, 904, 885, 856, 841, 835, 779, 759, 708, 641, 562, 481, 457; ¹H NMR (500 Hz, CDCl₃) δ (ppm) 0.90–0.98 (m, 1H), 1.35 (s, 9H), 1.94–2.00 (m, 2H), 2.33–2.41 (m, 1H), 2.78–2.87 (m, 2H), 3.93 (s, 3H), 6.98 (bs, 2H), 7.85 (d, *J* = 8.0 Hz, 1H); ¹³C NMR (125 Hz, CDCl₃) δ 20.9, 27.8, 35,0, 35.2, 40.2, 41.6, 56.7, 82.8, 115.8, 122.6, 125.9, 138.9, 140.5, 153.0, 168.6, 212.4, Anal. Calcd for C₁₈H₂₁NO₆: C, 62.24; H, 6.09; N, 4.03. Found: C, 62.17; H, 6.12; N, 4.09; MS (ESI, MeOH): 370 [M + Na]⁺.

6a. Yield 283 mg, 85%, ratio of stereoisomers = 1.9:1, separated by column chromatography.

6a'. Pale yellow oil, FT-IR (film, ν_{max}/cm^{-1}) 2981, 2244, 1721, 1597, 1562, 1520, 1471, 1392, 1368, 1351, 1277, 1248, 1155, 1125, 1077, 1025, 975, 950, 907, 864, 839, 786, 767, 729, 701; ¹H NMR (500 Hz, CDCl₃) δ (ppm) 1.42 (s, 9H, tBu), 1.54 (t, *J* = 7.0 Hz, 3H, CH₃CH₂), 1.98 (dd, *J* = 9.4 Hz, *J* = 5.0 Hz, 1H, CH₂), 2.34 (dd, *J* = 6.7 Hz, *J* = 5.0 Hz, 1H, CH₂), 2.56 (dd, *J* = 9.4 Hz, *J* = 6.7 Hz, 1H, CH-CN), 4.31 (q, *J* = 7.0 Hz, 2H, CH₃CH₂), 7.23 (s, 1H, ArH), 8.95 (s, 1H, ArH); ¹³C NMR (125 Hz, CDCl₃) δ 14.0, 14.2, 27.9, 37.4, 66.0, 83.9, 111.4, 117.2, 136.1, 146.3, 158.3, 158.4, 167.7; Anal. Calcd for C₁₆H₁₉N₃O₅: C, 57.65; H, 5.75; N, 12.61. Found: C, 57.50; H, 5.69; N, 12.73; MS (ESI, MeOH): 356 [M + Na]⁺.

6a". White solid, mp = 120–122 °C; FT-IR (KBr, $\nu_{\rm max}/\rm{cm}^{-1}$) 2984, 2249, 1712, 1603, 1562, 1523, 1488, 1472, 1455, 1394, 1365,1352, 1320, 1295, 1248, 1208, 1157, 1126, 1109, 1087, 1069, 1029, 972, 937, 869, 841, 771, 765, 730, 571, 505, 468, 440; ¹H NMR (500 Hz, CDCl₃) δ (ppm) 1.52–1.56 (m, 12H, 3H from <u>CH₃CH₂</u> and 9H from *t*Bu), 1.94 (dd, *J* = 9.0 Hz, *J* = 5.0 Hz, 1H, CH₂), 2.27 (dd, *J* = 7.0 Hz, *J* = 5.0 Hz, 1H, CH₂), 2.27 (dd, *J* = 7.0 Hz, *J* = 5.0 Hz, 1H, CH₂), 2.76 (dd, *J* = 9.0 Hz, *J* = 7.0 Hz, 1H, CH-CN), 4.30 (q, *J* = 7.0 Hz, 2H, CH₃CH₂), 7.56 (s, 1H, ArH), 8.81 (s, 1H, ArH); ¹³C NMR (125 Hz, CDCl₃) δ 14.2, 14.7, 22.5, 28.0, 36.6, 65.9, 84.4, 109.6, 117.4, 135.5, 146.2, 158.4, 159.9, 166.4; Anal. Calcd for C₁₆H₁₉N₃O₅: C, 57.65; H, 5.75; N, 12.61. Found: C, 57.62; H, 5.72; N, 12.50; MS (ESI, MeOH): 356 [M + Na]⁺.

6b'. Yield 275 mg, 72%, yellow oil; FT-IR (film, ν_{max}/cm^{-1}) 2980, 1717, 1596, 1561, 1520, 1471, 1392, 1367, 1352, 1279, 1256, 1246, 1200, 1155, 1126, 1085, 1027, 997, 961, 908, 857, 840, 803, 752, 727, 668; ¹H NMR (500 Hz, CDCl₃) δ (ppm) 1.11 (t, *J* = 7.1 Hz, 3H, CH₃-CH₂), 1.38 (s, 9H, *t*Bu), 1.50 (t, *J* = 7.0 Hz, 3H, CH₃-CH₂), 1.89 (dd, *J* = 8.5 Hz, *J* = 4.5 1H, Hz, CH₂), 2.09 (dd, *J* = 6.6 Hz, *J* = 4.5 Hz, 1H, CH₂), 2.76 (dd, *J* = 8.5 Hz, *J* = 6.6 Hz, 1H, COOEt), 4.20–4.29 (m, 2H, CH₃-CH₂), 6.98 (s, 1H, ArH), 8.90 (s, 1H, ArH); ¹³C NMR (125 Hz, CDCl₃) δ 14.0, 14.1, 19.8, 27.8, 29.0, 39.6, 61.2, 65.8, 82.8, 111.0, 135.5, 146.0, 158.3, 161.2, 169.1, 169.3; Anal. Calcd for C₁₈H₂₄N₂O₇: C, 56.83; H, 6.36; N, 7.36. Found: C, 56.72; H, 6.45; N, 7.20; MS (ESI, MeOH): 403 [M + Na]⁺, 783 [2M + Na]⁺.

6c. Yield 245 mg, 54%, yellow oil; FT-IR (film, ν_{max}/cm^{-1}) 2981, 1725, 1597, 1562, 1524, 1470, 1391, 1368, 1352, 1334, 1279, 1246, 1206, 1149, 1118, 1095, 1026, 986, 933, 859, 838, 766, 750, 726, 668; ¹H NMR (500 Hz, C_6D_6) δ (ppm) 0.75 (t, J = 7.0 Hz, 3H, $COOCH_2CH_3$), 0.82 (t, J = 7.0 Hz, 3H, OCH_2CH_3), 0.96 (t, J = 7.0Hz, 3H, COOCH2CH3), 1.34 (s, 9H, tBu), 3.07-3.20 (m, 2H, OCH_2CH_3 , 3.57 (d, J = 6.5 Hz, 1H, CH-COOEt), 3.69 (d, J = 6.5Hz, 1H, CH-COOEt), 3.73 (dd, J = 14 Hz, J = 7.0 Hz, 2H, COOCH2CH3), 3.94-4.01 (m, 2H, COOCH2CH3), 6.91 (s, 1H, ArH), 8.74 (s, 1H, ArH); ¹H NMR (500 Hz, CDCl₃) δ (ppm)1.13 (t, J = 7.1 Hz, 3H, CH_2CH_3), 1.32 (t, J = 7.1 Hz, 3H, CH_2CH_3), 1.39 (s, 9H, tBu), 1.51 (t, J = 7.0 Hz, 3H, <u>CH₃CH₂</u>), 3.26 (s, 2H, <u>CH</u>COOEt), 2.00 (m, 2H, CH₃CH₂), 4.25 (m, 4H, CH₂CH₃), 7.04 (s, 1H, ArH), 8.88 (s, 1H, ArH); ${}^{13}C$ NMR (125 Hz, C_6D_6) δ 13.8, 13.8, 14.1, 27.7, 32,6, 33.1, 47.2, 61.4, 61.7, 65.3, 82.8, 110.0, 136.5, 146.2, 158.5, 160.0, 166.0, 167.6, 168.0; ¹³C NMR (125 Hz, CDCl₃); δ 14.0, 14.1, 14.2, 27.7, 32,2, 32.3, 46.3, 61.6, 61.8, 66.0, 83.4, 109.7, 135.8, 146.2, 158.5, 159.80, 165.6, 167.6, 167.7; Anal. Calcd for $C_{21}H_{28}N_2O_9$: C, 55.75; H, 6.24; N, 6.19. Found: C, 55.60; H, 6.14; N, 5.99; MS (ESI, MeOH): $475 [M + Na]^+$

Reaction of Ethyl Chloro(2-nitro-5-chlorophenyl)acetate with Acrylonitrile under Solid–Liquid PTC Condition. To a suspension of anhydrous potassium carbonate (1382 mg, 10 mmol) in dry DMF (10 mL), solid $Bu_4N^+HSO_4^-$ (34 mg, 0.1 mmol) was added, at room temperature. The mixture was intensively stirred for 10 min, and a solution of ethyl chloro(2-nitro-5-chlorophenyl)acetate (1 mmol) and acrylonitrile (1.5 mmol) in 5 mL of DMF was added. The mixture was stirred at room temperature until aryl ester was consumed (TLC control). The reaction mixture was poured in water (20 mL), and the product was extracted with dichloromethane (3 × 15 mL). The combined extract was washed with water (3 × 20 mL) and dried, and the solvent was evaporated. The products were purified by column chromatography on silica gel, hexane–ethyl acetate eluent. Evapo-

ration of the solvent gave final products as single or a mixture of stereoisomers.

7a. Yield 24 mg, 8%, white solid; mp = 129.5–131.5 °C; ¹H NMR (500 Hz, CDCl₃) δ (ppm) 1.22 (t, *J* = 7.0 Hz, 3H, CH₃), 1.68 (bs, 1H, CH₂), 2.18–2.21 (m, 1H, CH₂), 2.42–2.44 (m, 1H, CH-CN), 4.24 (q, *J* = 7.0 Hz, 2H, <u>CH₂</u>CH₃), 7.45 (d, *J* = 1.5 Hz, 1H, ArH), 7.53 (dd, *J* = 8.5 Hz, *J* = 1.5 Hz, 1H, ArH), 8.10 (d, *J* = 8.5 Hz, 1H, ArH); ¹³C NMR (125 Hz, CDCl₃) δ 13.9, 14.9, 20.9, 34.5, 62.6, 116.5, 127.0, 129.9, 132.3, 133.5, 140.2, 147.5, 167.1; Anal. Calcd for C₁₃H₁₁ClN₂O₄: C, 52.98; H, 3.76; Cl, 12.03; N, 9.51. Found: C, 52.86; H, 3.84; Cl, 12.07; N, 9.46; MS (ESI, MeOH): 317 [M + Na]⁺, 611 [2M + Na]⁺.

7aa. Yield 37 mg, 13%, pale yellow solid, mp = 157-160,5 °C; ¹H NMR (500 Hz, CDCl₃) δ (ppm) 1.48 (t, J = 7.1 Hz, 3H, CH₃), 4.50 (q, J = 7.1 Hz, 2H, CH₂), 7.77 (dd, J = 9.3 Hz, J = 1.8 Hz, 1H, ArH), 8.27 (s, 1H, ArH), 8.63 (d, J = 9.3 Hz, 1H, ArH), 9.20 (d, J = 1.7 Hz, 1H, ArH); ¹³C NMR (125 Hz, CDCl₃) δ 14.2, 62.6, 112.0, 120.7, 121.5, 121.7, 126.5, 127.9, 129.5, 132.4, 139.6, 141.2, 162.9; Anal. Calcd for C₁₃H₉ClN₂O₃: C, 56.43; H, 3.28; Cl, 12.81; N, 10.13. Found: C, 56.26; H, 3.13; Cl, 12.79; N, 9.98; MS (ESI, MeOH): 299 [M + Na]⁺, 575 [2M + Na]⁺.

ASSOCIATED CONTENT

S Supporting Information

¹H NMR chemical shifts and coupling constants of cyclopropane ring protons in tabular form, assignment of the stereochemistry of the cyclopropanes, and copies of the ¹H NMR and ¹³C NMR spectra of all new compounds.The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.joc.5b00204.

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Notes

The authors declare no competing financial interest.

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